

Effects of hypothalamic hormones (GRF, TRH, Somatostatin) and insulin-like growth factor I on growth hormone secretion from prepubertal male lamb pituitary cultures

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Summary. We have examined the regulation of GH secretion from monolayer cultures of prepubertal male lamb anterior pituitary cells. Growth hormone-releasing factor (GRF 1-44) stimulated GH release in a dose-related manner : the maximal effective dose was 10^{-10} M, which caused a 500 % increase in basal GH secretion, while the half-maximal effect was reached with a dose of 2.5×10^{-11} M (ED_{50}). Thyrotropin-releasing hormone (TRH) also elicited a dose-dependent stimulation of GH secretion, although it was approximately 1000 times less potent than GRF. GRF and TRH did not have additive or synergistic effects on GH secretion.

Somatostatin (SRIF) at a concentration of 10^{-7} M maximally inhibited basal GH release to 40 % of that of the control ; the ED_{50} was 2.0×10^{-9} M. Moreover, 10^{-7} M SRIF blocked the stimulation of GH secretion induced by 10^{-8} M GRF. However, when the cells were incubated with these two peptides at an identical concentration (10^{-8} M), GH secretion was stimulated significantly above control values. When added at the same concentration (10^{-7} M), TRH and SRIF nullified their respective effects. A dose of 100 ng/ml of synthetic IGF-I was without effect on basal GH release, but significantly decreased 10^{-9} M GRF-induced stimulation of GH secretion.

These data indicate that in prepubertal male lambs : (1) the stimulatory effect of GRF is predominant over the inhibitory effect of SRIF, (2) somatostatin inhibits TRH stimulation of GH secretion *in vitro*, and (3) IGF-I may control GH secretion by modulating GRF effects at the pituitary level.

Introduction.

Two hypothalamic hormones, growth hormone-releasing factor (GRF) and growth hormone release-inhibiting factor (SRIF), are known to be of major importance in regulating growth hormone (GH) secretion in a variety of animal

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species, including man. In normal humans, thyrotropin-releasing hormone (TRH) regulates TSH secretion but does not modify GH secretion (Anderson *et al.*, 1971). A stimulatory effect of TRH on GH secretion *in vivo* and *in vitro* has been shown in chickens (Leung and Taylor, 1983), acromegalic patients (Jialal *et al.*, 1984), and hypothyroid rats (Szabo *et al.*, 1984). Moreover, a synergistic interaction between TRH and GRF on GH secretion has recently been demonstrated in immature rat pituitary cultures (Welsh *et al.*, 1986) and in cattle *in vivo* (Hodate *et al.*, 1985) and *in vitro* (Ingram and Bicknell, 1986). These results suggest that, under certain conditions, TRH is also able to modulate GH secretion. In addition, numerous data support the concept that in humans and rats, insulin-like growth factors (IGFs) regulate GH secretion at the pituitary level by a negative feedback mechanism (Brazeau *et al.*, 1982a ; Tannenbaum *et al.*, 1983 ; Goodyer *et al.*, 1984a ; Goodyer *et al.*, 1986).

We have recently developed an ovine pituitary monolayer culture system in order to study the ontogeny of mechanisms which control GH secretion during fetal and post-natal life in sheep. In the present study, we have investigated the roles of GRF, TRH, SRIF and IGF-I, alone or in combination, in the regulation of GH secretion from prepubertal male lamb pituitary cells.

Materials and methods.

Materials. — Human synthetic somatotrin, GRF 1-44 (SR 95228), and somatostatin, SRIF 1-14 (SR 9357), were provided by Sanofi (Montpellier, France). Recombinant human (Thr59)-IGF-I was purchased from Amersham (Buckinghamshire, England). Collagenase, hyaluronidase, deoxyribonuclease, Ham's F-10 culture medium, fetal bovine serum and TRH were all obtained from Boehringer Laboratories (Mannheim, Germany). Egg albumin and bovine serum albumin were purchased from Sigma (St Louis, MO).

Preparation of cells for monolayer culture. — The pituitary glands of four prepubertal male « Mérimos d'Arles » lambs, removed immediately after slaughter, were placed in Ham's F-10 with antibiotics and processed individually for monolayer culture. The anterior lobes were separated from the posterior and intermediate lobes and minced with sterile scissors. The pituitary fragments were dispersed at 37 °C for 45 to 60 min in Ham's F-10 medium containing 1.5 mg/ml of collagenase, 0.5 mg/ml of hyaluronidase, 0.25 mg/ml of deoxyribonuclease and 3 % egg albumin. Cell dispersion was augmented by triturating the fragments through a sterile Pasteur pipette. Once the dispersion was completed, the cells were harvested by centrifugation for 2 min at 400 × g and washed twice in Ham's F-10 medium containing 10 % fetal bovine serum. The cells were then plated at a density of 50,000 cells/well in 12-well plastic tissue culture plates in Ham's F-10 supplemented with 10 % fetal bovine serum and antibiotics (5 µg/ml of amphotericin B, 200 IU/ml of penicillin G, 40 µg/ml of gentamycin sulfate). Cell cultures were incubated at 37 °C in a humidified atmosphere with 5 % CO₂ for 3 days before the experiments began. The media were changed daily after the first 48 h.

Experimental procedure. — Experimental incubations were performed on the fourth day of culture using Ham's F-10 culture medium supplemented with 1 % bovine serum albumin. The cells were preincubated for one hour, then rinsed and incubated for 3 h in the presence or absence of the test factors : each dose was tested in duplicate. At the end of the 3-hour incubation period, culture media were collected and centrifuged for 2 min at $400 \times g$ to remove any cellular material. The supernatants were kept at -80°C until assayed for oGH.

Ovine growth hormone radioimmunoassay. — Ovine GH was measured by specific radioimmunoassay using a double antibody separation method. Reagents were provided by the National Hormone and Pituitary Program (NIADDK, Bethesda, MD) with the exception of sheep anti-rabbit-gammaglobulin serum which was prepared in our laboratory. The sensitivity of the assay was 0.5 ng/ml and the intra-assay variation was 5 %. Appropriate samples were prescreened and then all samples were run in the same assay at the appropriate dilution.

Expression of the results. — Results were expressed as a percentage of the controls (mean \pm SE). Data were statistically analysed using analysis of variance followed by Duncan's multiple-range test after logarithmic transformation. The ED_{50} s were calculated as described by Rodbard (1974).

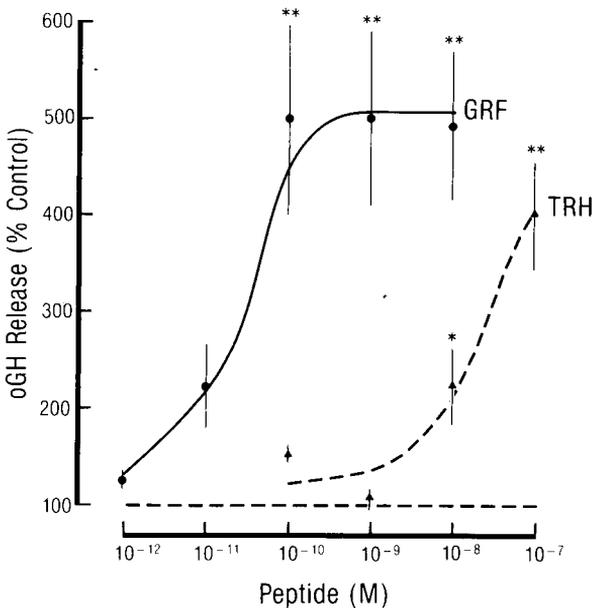


FIG. 1. — Dose-response curve of the effect of GRF or TRH on oGH release from cultured male lamb pituitary cells. Data are expressed as percent of control values (mean \pm SE ; n = 4) * $p < 0.05$, ** $p < 0.01$: statistical significance as compared to control values (100 % = 9.31 ± 1.54 ng/ml/3 h).

Results.

Effect of GRF and TRH on GH release. — Basal GH secretion from cultured lamb pituitary cells was 9.31 ± 1.54 ng/ml/3 h. The cells responded in a dose-related fashion to GRF 1-44 (10^{-12} M- 10^{-8} M) (fig. 1). Maximal stimulation occurred with a dose of 10^{-10} M, corresponding to an increase of 500 % of basal GH secretion ($p < 0.01$). The half-maximal effect was obtained with a dose of 2.5×10^{-11} M (ED_{50}).

TRH also stimulated oGH release in a dose-dependent manner : the lowest effective dose ($p < 0.05$) was 10^{-8} M, and a concentration of 10^{-7} M caused a 400 % increase in basal secretion ($p < 0.01$). The ED_{50} for TRH could not be determined since a maximal stimulation was not achieved. However, GRF was approximately 1,000 times more potent than TRH in stimulating GH release since the same effect was achieved with 10^{-11} M GRF or 10^{-8} M TRH.

Effect of somatostatin on GH release. — Somatostatin (10^{-10} M- 10^{-6} M) caused a dose-related inhibition of GH secretion (fig. 2). Hormone output was maximally decreased to 40 % of its basal value with 10^{-7} M SRIF ($p < 0.01$) : the ED_{50} was 2.0×10^{-9} M.

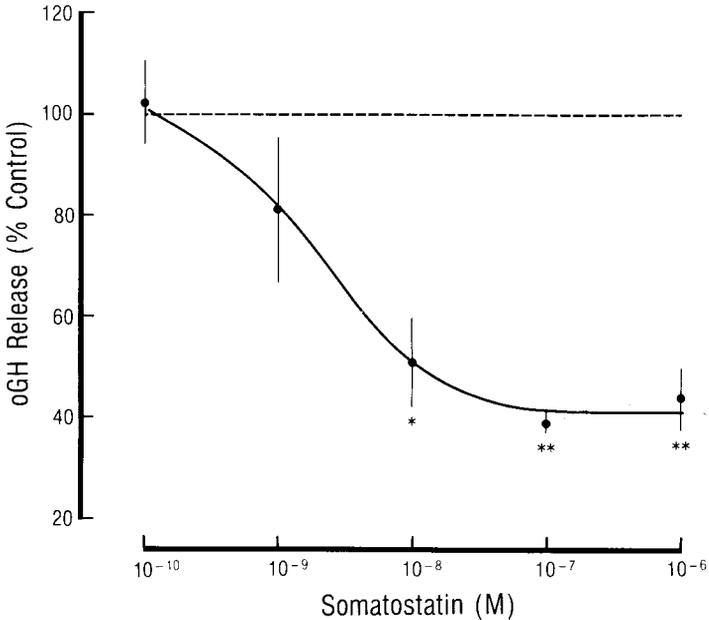


FIG. 2. — Dose-response curve of the effect of SRIF on oGH release from cultured male lamb-pituitary cells. Data are expressed as percent of control values (mean \pm SE ; $n = 4$) * $p < 0.05$, ** $p < 0.01$: statistical significance as compared to control values (100 % = 9.31 ± 1.54 ng/ml/3 h).

Effect of GRF 1-44 plus somatostatin. — Figure 3 represents two different experiments showing the effect of combining GRF and somatostatin. In the first experiment (fig. 3A), increasing doses of somatostatin (10^{-10} M- 10^{-6} M) were

tested in the presence of a constant concentration of 10^{-8} M GRF. A 10^{-7} M dose of SRIF completely abolished GRF stimulation of GH secretion (from 492.2 ± 84.9 % to 95.5 ± 19.5 % ; $p < 0.01$). When GRF and somatostatin were added to the cells at the same concentration (10^{-8} M), the resulting effect was a significant ($p < 0.01$) increase above basal GH secretion, indicating that the stimulatory effect of GRF on somatotrope activity was predominant over the inhibitory effect of SRIF in prepubertal lambs. In the second experiment (fig. 3B), a constant dose of 10^{-7} M SRIF was added in combination with increasing concentrations of GRF ranging from 10^{-11} to 10^{-8} M. GRF blocked the inhibition induced by somatostatin but did not significantly increase oGH release above control values.

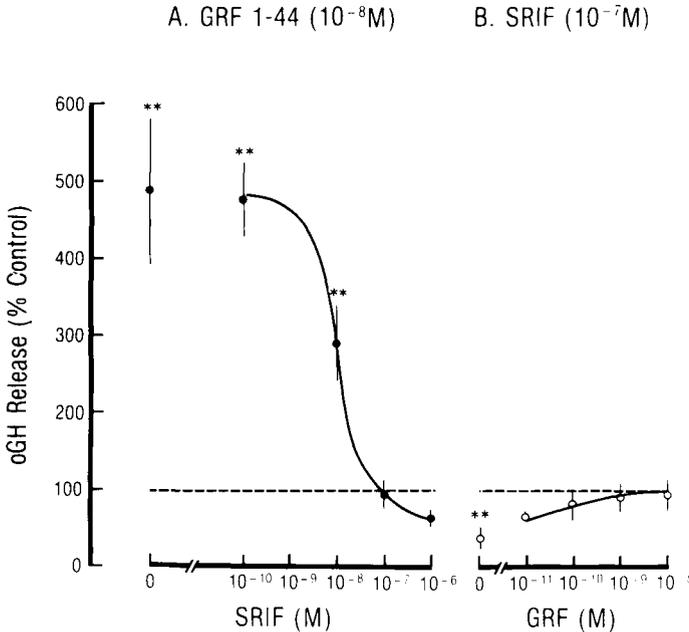


FIG. 3.— Interactions of GRF and SRIF on oGH release from male lamb pituitary cells in culture : A. Effect of GRF (10^{-8} M) on SRIF (10^{-10} M- 10^{-6} M) inhibition of GH release, and B. Effect of SRIF (10^{-7} M) on GRF (10^{-11} M- 10^{-8} M) stimulation of GH release.

Data are expressed as percent of control values (mean \pm SE ; $n = 4$) ; ** $p < 0.01$: statistical significance as compared to control values (100 % = 9.31 ± 1.54 ng/ml/3 h).

Effect of TRH plus GRF or somatostatin. — A constant dose of TRH (10^{-7} M) was tested in the presence of increasing concentrations of GRF (10^{-12} M- 10^{-8} M) or somatostatin (10^{-11} M- 10^{-7} M) (fig. 4). GRF did not significantly increase TRH stimulation of GH secretion. Furthermore, the response induced by 10^{-8} M GRF plus 10^{-7} M TRH was not significantly different from the effect of 10^{-8} GRF alone. Thus, we did not observe any additive or synergistic effects of GRF and TRH on oGH secretion.

TRH-induced stimulation of GH release was markedly inhibited by SRIF (from 402.6 ± 62.0 % to 92.7 ± 2.8 % ; $p < 0.01$). In contrast to what we observed

with GRF and somatostatin, incubation of cells with the same dose of TRH and SRIF (10^{-7} M) did not modify basal GH secretion. These results indicate that, in prepubertal lambs, SRIF is able to completely block TRH stimulation of GH release at the pituitary level.

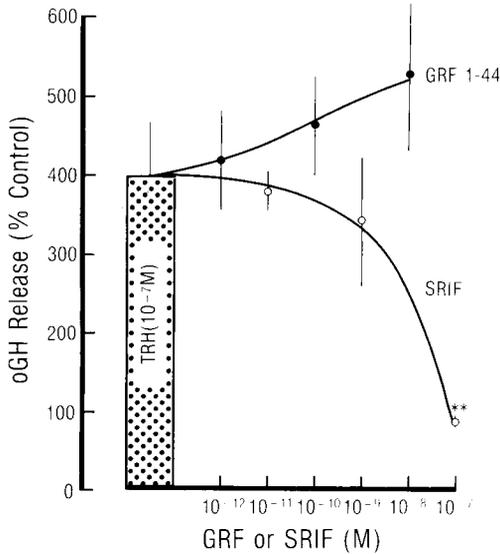


FIG. 4. — Interactions of TRH (10^{-7} M) with GRF (10^{-12} M – 10^{-8} M) or SRIF (10^{-11} M – 10^{-7} M) on GH secretion from cultured male lamb pituitary cells. Data are expressed as percent of control values (mean \pm SE; n = 4) ** p < 0.01 : statistical significance as compared to 10^{-7} M TRH (100 % = 9.31 ± 1.54 ng/ml/3 h).

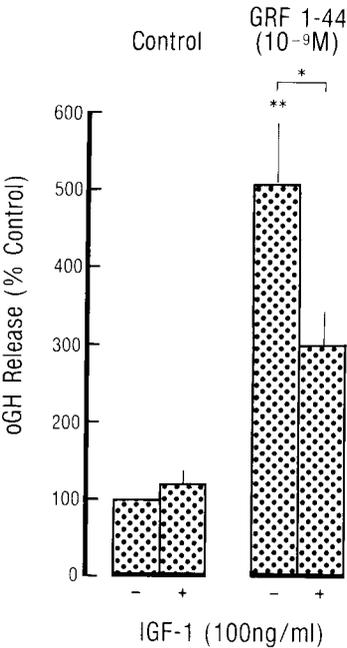


FIG. 5. — Effect of IGF-I (100 ng/ml) on basal and GRF (10^{-9} M) – stimulated GH secretion. Data are expressed as percent of control values (mean \pm SE; n = 4) ** p < 0.01 : statistical significance as compared to control values (100 % = 10.6 ± 1.97 ng/ml/3 h).

Effect of IGF-I on basal and GRF-stimulated GH secretion. — Figure 5 shows that while 100 ng/ml of synthetic IGF-I did not modify basal GH secretion during the three-hour incubation period, GRF (10^{-9} M)-stimulated release of GH was significantly decreased (from 508.04 ± 82.05 % to 299.68 ± 46.23 % ; $p < 0.05$).

Discussion.

Our results indicating that GRF is a potent stimulator of GH release from cultured pituitary cells of prepubertal male lambs are in agreement with *in vitro* pituitary data on lambs from Law *et al.* (1984). They also correlate with the finding that GRF stimulates GH release in prepubertal male lambs *in vivo* (Barenton *et al.*, 1987). The GRF dose-response curve for these cells is similar to that observed in studies using adult male rat pituitary cultures (Brazeau *et al.*, 1982b).

We also found that somatostatin can inhibit basal as well as GRF-stimulated GH secretion, although the cells appear to be more sensitive to GRF than to SRIF: that is, in prepubertal lambs, the effect of GRF on GH secretion is predominant over the effect of SRIF when the same concentrations are tested. A similar phenomenon has been reported in studies using fetal human pituitary explants (Goodyer *et al.*, 1984b; Goodyer *et al.*, 1987), while the opposite phenomenon has been observed with adult rat pituitary cultures (Law *et al.*, 1985). These discrepancies may be due to species differences or to changes in the relative potencies of GRF and SRIF during development.

We have also clearly shown that TRH stimulates GH release from lamb pituitary cell cultures, even though it is much less effective than GRF. It is unlikely that TRH acts through the GRF receptor since the respective structures of the two peptides are so different. On the other hand, the existence of two different populations of specific receptors for GRF and TRH on the somatotrophs (as shown for human pituitary adenomas: Le Dafniet *et al.*, 1985) would imply that submaximal doses of GRF and TRH have additive effects on GH secretion. We did not observe any additive or synergistic effects with the two peptides, but the elevated dose of TRH (10^{-7} M) we used in these experiments may have masked certain subtle interactions. These data suggest that a common final pathway may be involved in GRF and TRH action mechanisms. Further experiments will be required to elucidate the physiologic significance of the action of TRH on GH secretion in lambs.

We have also demonstrated that SRIF is able to abolish TRH-induced stimulation of GH release *in vitro*. It is noteworthy that, in contrast to what we observed with GRF and SRIF, the addition of equivalent doses of TRH and SRIF to the cells resulted in a complete blockade of the effect of TRH. Wrutniak *et al.* (1987) did not observe any *in vivo* effect of TRH on GH secretion in lambs. These findings suggest that, *in vivo*, endogenous somatostatin might inhibit response to TRH by blocking its effect at the pituitary level.

Growth hormone has no single well-defined target organ, but has been shown to induce the hepatic production of somatomedins (Uthne and Uthne,

1972 ; Schimpff *et al.*, 1976 ; Binoux *et al.*, 1980) which may mediate the peripheral effects of GH (Froesch *et al.*, 1985). Clinical and experimental observations have led to the concept of a role for the somatomedins in controlling GH secretion by a negative feedback loop on the pituitary and hypothalamus. Low levels of IGFs are often associated with high levels of GH (Zapf *et al.*, 1976). IGFs suppress the episodic bursts of GH when injected intraventricularly into freely-moving rats (Tannenbaum *et al.*, 1983). Recently, Yamashita and Melmed (1986) showed that IGF-I inhibits GH synthesis by decreasing GH mRNA levels.

We did not see any effect of IGF-I on basal GH secretion in our experiment, although we used a pure peptide in a defined medium. Goodyer *et al.* (1984a ; 1986) reported an effect of 100 ng/ml of purified IGF-I on basal GH secretion from adult rat and fetal human pituitary explants after 3-4 h while Brazeau *et al.* (1982a) observed the same effect at a lower dose but after 20 hours of incubation. Results may differ, depending on the species, dose, length of incubation or the origin of the peptide used.

In contrast, IGF-I decreased GRF-induced stimulation of GH release. These data are consistent with previous observations with rat pituitary monolayers (Brazeau *et al.*, 1982a). They support the hypothesis that IGF-I can exert a negative feedback inhibition of oGH synthesis and/or release by modulating the effect of GRF on somatotrope activity in prepubertal lambs. Further studies will be necessary to determine the role for somatomedins during fetal and post-natal development in sheep.

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Résumé. *Effet des hormones hypothalamiques (GRF, TRH, somatostatine) et du facteur de croissance (IGF-I) sur la sécrétion de la GH par des cultures d'hypophyse d'agneau mâle prépubère.*

Nous avons étudié la régulation de la sécrétion de l'hormone de croissance par les cellules hypophysaires ovines en culture primaire. Le GRF 1-44 stimule la libération de la GH de façon dose-dépendante : l'effet maximum est atteint avec une concentration de 10^{-10} M de GRF, et représente une augmentation de 500 % de la sécrétion basale. La dose qui induit 50 % de l'effet maximum (ED_{50}) est de $2,5 \times 10^{-11}$ M. La thyroélibérine (TRH) entraîne également une stimulation de la sécrétion de GH, bien que la sensibilité des cellules vis-à-vis de la TRH soit environ 1 000 fois plus faible que celle vis-à-vis du GRF. Les effets du GRF et de la TRH sur la sécrétion de GH ne sont ni additifs, ni synergiques. La somatostatine inhibe la libération basale de GH de 60 % à la dose de 10^{-7} M : l' ED_{50} est de $2,0 \times 10^{-9}$ M. De plus, la stimulation de la sécrétion de GH induite par 10^{-8} M de GRF est bloquée par 10^{-7} M de somatostatine. Cependant, quand on incube les cellules en présence de ces deux peptides à concentration égale (10^{-8} M), l'effet résultant observé est une stimulation de la libération de GH. Par ailleurs, les effets de la TRH et du SRIF s'annulent quand les cellules sont incubées avec les deux peptides à la même concentration. Enfin, une dose de 100 ng/ml d'IGF-I synthétique est sans effet sur la libération basale de GH, mais diminue la stimulation induite par 10^{-9} M de GRF.

Ces résultats indiquent que chez l'agneau mâle prépubère : 1) l'effet stimulant du GRF prédomine sur l'effet inhibiteur de la somatostatine ; 2) la somatostatine inhibe la stimulation de la sécrétion de GH induite par la TRH *in vitro* ; et 3) IGF-I pourrait participer au contrôle de la sécrétion de la GH, en modulant l'action du GRF au niveau de l'hypophyse.

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