

NOTE

RETENTION OF NUCLEAR BASIC PROTEIN
IN RABBIT SPERMATOZOA UP TO ENTRY
INTO THE VITELLUS

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The dynamism of basic nuclear protein has been studied in mammalian male germ cells in mouse, bull and ram up to the epididymal spermatozoon stage (MONESI, 1964, 1965; GLEDHILL *et al.*, 1966; LOIR, 1972 *a* and *b*). After natural mating in mouse, we find that arginine ¹⁴C-labelled nuclear protein is retained in ejaculated spermatozoa in *ampullae oviducti* in the close vicinity of the ovum (KOPEČNÝ, 1970). However, KOPEČNÝ and PAVLOK (1975), studying ova fertilized *in vitro*, concluded that this protein is suddenly lost during the earliest stages of male pronuclear formation. The particularly rapid disappearance of labelled nuclear protein in fertilization led us to reinvestigate the presumed resistance of similar labelling in spermatozoa of another species during independent life in the female genital tract. Rabbit seemed especially suited to this purpose because spermatozoa must spend a much longer time in the female genital tract before entering the ovum in this species than in the mouse. Some observations indicate a change in rabbit sperm chromatin during its passage through the female genital tract (CHANG, 1959; ORGBIN-CRIST, 1969). In addition to this, supplementary spermatozoa accumulate in considerable quantities in the perivitelline space of rabbit oocyte.

Four mature male rabbits were injected intratesticularly as described elsewhere (FULKA *et al.*, 1967) with radioactive arginine. Males n^o 1-3 received 150 µCi of L-arginine-5-³H-mono-hydrochloride, specific activity 500 mCi/mmol (Amersham), in each testis on day 0; male n^o 4 was similarly injected with 25 µCi L-arginine-¹⁴C (U), specific activity 160 mCi/mmol (U. V. V. R., Prague), but the injections were repeated once more two days later.

Rabbit n^o 1 was hemicastrated after 24 hours and 4 days p.i., and ejaculates were collected from rabbits 2, 3 and 4 twice weekly. Sperm samples were further isolated from the ductus deferens and caudae epididymidis on the day of the last collection (fig. 1). Four does were inseminated after HCG injection (100 IU) with the ejaculate from male n^o 4, collected on day 17 p.i. Fourteen hours after insemination, these females were slaughtered, and the eggs isolated, mounted on slides, air-dried, and subsequently fixed with acetic alcohol. Histological sections from the testes, and smears of ejaculated and isolated spermatozoa and egg preparations were coated

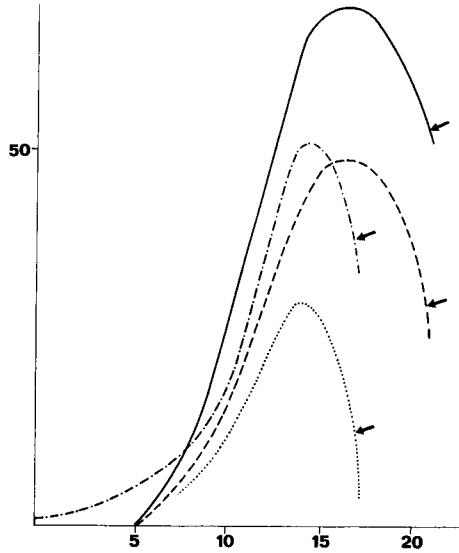


FIG. 1. — Availability of labelled rabbit spermatozoa in ejaculates (all curves up to the point marked by an arrow) or in excised ductus deferens and caudae epididymidis (after arrow) following intratesticular injection of ^3H - or ^{14}C -arginine. Abscissa : days after 1st injection. Ordinate : per cent of labelled spermatozoa.

Males n° 2 and 3 (mean)	{	—————	4 and more grains/sperm head
^3H -arginine, injection day 0		- - - - -	10 and more grains/sperm head
Male n° 4. ^{14}C -arginine,	{	- · - · - ·	4 and more grains/sperm head
injections on days 0 and 2		· · · · ·	30 and more grains/sperm head

Sperm was collected twice weekly. On the day of the last collection (males n° 2 and 3 : 21st day p.i., male n° 4 : 17th day p.i.) ductus deferens, proximal and distal caudae epididymidis were excised and spermatozoa isolated from them, respectively. The labelling data from all cells were included in the graphs.

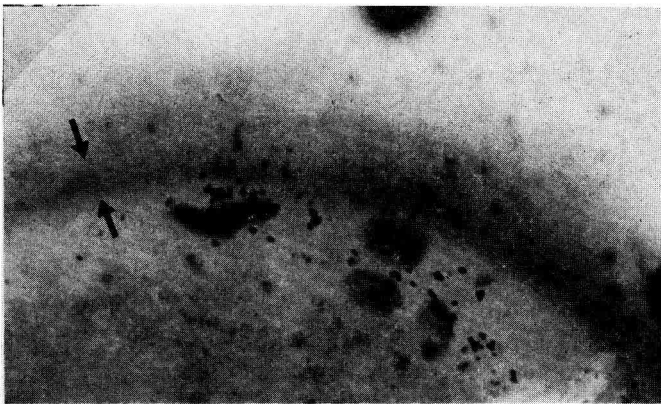


FIG. 2. — Autoradiogram of supplementary rabbit spermatozoa, labelled with ^{14}C -arginine, in perivitelline space of a fertilized oocyte

Thick layer of nuclear emulsion Ilford K5, exposure 119 days.
Most of length of the ^{14}C -tracks originating in spermatozoa is out of focus.
Between arrows : zona pellucida

with nuclear emulsion Ilford K 5 and exposed for varying lengths of time. The oocyte preparations were exposed for 119 days and stained with toluidine blue (Gurr) at pH 4 after development.

Intensive labelling was found in elongating spermatid nuclei in testis. A wave of cells in ejaculated spermatozoa showed intensive nuclear labelling with a peak at about 15 days p.i. (fig. 1). In fourteen fertilized oocytes (two pronuclei) many supplementary spermatozoa were observed to be the origin of carbon-14 tracks (fig. 2). No label was found in early pronuclei.

As observed in this material, the arginine-³H incorporation in elongation rabbit spermatids probably parallels a similar situation in mouse (MONEST, 1964, 1965) and in ram (LOIR, 1972 a); it may be considered as localization of the synthesis of basic nuclear arginine-rich spermatozoan protein in these cells. Although repeated intraperitoneal injections in the mouse lead to general spermatozoal labelling (KOPĚČNÝ and PAVLOK, 1975), sufficient labelling of ejaculate spermatozoa is not achieved by introducing the precursor into the testis via one or two successive intratesticular injections in the rabbit. Thus, total absence of the label in early pronuclei in rabbit cannot be interpreted as a change in nuclear spermatozoal protein during fertilization. On the other hand, the presence of supplementary labelled spermatozoa in the perivitelline space is convincing proof for retention of most of the basic nuclear protein *in situ*, even after the spermatozoa have undergone capacitation and acrosomal reaction in the female genital tract and penetrated through the zona pellucida. This confirms that the spermatozoal basic nuclear protein of mammals, rich in arginine and synthesized in elongating spermatids, is stable during the independent life of mammalian sperm and does not undergo the transformations observed in some non-mammalian species (MONEST, 1971). The quantitative change in this protein is detectable only during sperm head swelling in the ooplasm (KOPĚČNÝ and PAVLOK, 1975). Retention of the label in spermatozoa does not relate, of course, to possible biochemical modifications of spermatozoan basic protein, which would not affect its arginine content.

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RÉSUMÉ

LA RÉTENTION DE L'ARGININE-¹⁴C DES NUCLÉOPROTÉINES DES SPERMATOZOÏDES DE LAPIN JUSQU'À LA PÉNÉTRATION DANS LE VITELLUS

Le marquage par l'arginine-¹⁴C des nucléoprotéines des spermatozoïdes de Lapin au cours de la spermiogénèse, permet de suivre leur évolution jusqu'à la fécondation.

Aucune modification ne se produit pendant le séjour dans les voies génitales femelles, la capacitation et le franchissement de la membrane pellucide. L'absence de radioactivité dans le pronucleus mâle au début de sa formation laisse penser que ces nucléoprotéines se transforment dès la pénétration du spermatozoïde dans le cytoplasme de l'ovocyte, comme les auteurs l'ont montré d'une manière indiscutable chez la Souris.

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